# *In vitro* Evaluation (Antioxidant Activity) of coumarin derivatives

Devi Sarika\*, Singh Pooja, Rathi Vaibhav and Kumar Praveen Quantum School of Health Science, Quantum University, Roorkee, 247667, INDIA \*Sarikasaini1991@gmail.com

### Abstract

Coumarin is a heterocyclic molecule associated with beneficial human health effects such as to reduce the risk of cancer, diabetes, cardiovascular and brain disease. These effects are thought to be related to free radical scavenging due to their antioxidant properties. Coumarin is a substance that has been synthesized in many of its derivatives in recent years. This entity is a major source of interest for many medicinal chemists to explore its various pharmacological possibilities, especially anticoagulant activity.

The antioxidant activities of two synthesized coumarin derivatives namely 2-(4-methyl-2-oxo-2H-chromen-7yloxy)-N'-(1-(4-aminophenyl)ethylidene) acetohydra zide [M1] and 2-(4-methyl-2-oxo-2H-chromen-7yloxy)-N'-(1-(2 hydroxy phenyl) ethylidene) acetohydrazide [M2] were studied with the DPPH method. Structure of two coumarin synthesized compounds is proposed on the basis of spectroscopic evidence.

**Keywords:** Coumarin, Antioxidant, DPPH, Synthesis, Ascorbic acid.

# Introduction

Coumarin is one of the potent secondary metabolites of plants<sup>1,2</sup> and fungi<sup>3</sup> and is characterized by multiple pharmacological properties<sup>4</sup>. Like declucin and declucinol, these coumarins have a pyranocoumarin moiety isolated from the medicinal plant Angelica<sup>5</sup>. Many of these compounds are antibacterial<sup>6-8</sup>, antifungal<sup>9</sup>, anti-inflammatory<sup>10</sup>, anticoagulant<sup>11</sup>, anti-HIV<sup>12</sup> and antitumor<sup>13</sup>.

It is shown that coumarin is commonly used as an additive in foods, perfumes, cosmetics<sup>14</sup>, pharmaceuticals and optical brighteners<sup>15</sup> and disperses fluorescent and laser dyes<sup>16</sup>. Coumarin also has excellent optical properties such as superthermal stability, extended spectral sensitivity, high quantum yield and excellent optical stability<sup>17</sup>.

The anti-inflammatory activity of coumarin-derived compounds has been extensively reviewed and a structureactivity relationship (SAR) has been established with antiinflammatory effects when aromatic groups are directly fused at the 3-position of the coumarin nucleus or bound via intermediate bonds induces. Many of these derivatives also have antioxidant activity due to the scavenging mechanism<sup>18</sup>. A coumarin agent consisting of a fused ring of benzene and pyrone (known as 1,2-benzopyrone) is abundant in plants and more than 1300 coumarins have been identified from natural resources<sup>19</sup>.

The synthesis of coumarins and their derivatives has received considerable attention from organic and medicinal chemists over the years, as many natural products contain this heterocyclic core<sup>20</sup>.

Therefore, the synthesis of this heterocyclic core is very interesting. Coumarin has been synthesized by several routes including the Pechmann<sup>21</sup>, Parkin<sup>22</sup>, Knoevenagel<sup>23</sup>, Reformatsky<sup>24</sup> and Wittig<sup>25</sup> reactions. Coumarin also has anticoagulant properties, some of which are commonly used as anticoagulants such as warfarin and acenocoumarol<sup>26-29</sup>.

Antioxidants have the ability to protect organelles from damage caused by oxidative stress caused by free radicals. The free radicals used include hydroxyl radicals, superoxide anion radicals and hydrogen peroxide. Reactive free radicals formed by exogenous chemicals, stress, or food systems can oxidize biomolecules, causing cancer, coronary artery disease and hypertension<sup>30</sup>.

In general, most of the metabolically produced free radicals are removed by endogenous defense systems such as catalase, superoxide dismutase and the peroxidase-glutathione system<sup>31</sup>.

# **Material and Methods**

**Materials:** FTIR spectra of the samples are performed by FTIR Spectrophotometer MODEL-8300 of SHIMADZU, in the region of 400 4000 cm<sup>-1</sup>. Melting points were recorded by the melting point apparatus. All reactions and product purity were examined by thin layer chromatography (TLC) on aluminum back panels coated with silica gel.

All chemicals were purchased from Aldrich Sigma Company. NMR spectra of the synthesized compounds were recorded in DMSO (dimethyl sulfoxide) using BROKE JEOL Model AV300 at 300 MHz spectrometer.

**Synthesis of 2-(4-methyl-2-oxo-2H-chromen-7-yloxy)-N'-**(**1-(4-aminophenyl)-ethylidene)-acetohydrazide** [**M1**]<sup>32</sup>: A mixture of 2-3 drops of glacial acetic acid in compound 2-(4-methyl-2-oxo-2H-chromen-7-yloxy) acetohydrazide (0.0008 mol), 4-aminoacetophenone (0.0008 mol) and anhydrous alcohol (60 mL) was refluxed on a water bath for 16 hours. The reaction mixture was poured into crushed ice. It is filtered, dried and recrystallized from ethanol to give compound (M1).



Scheme 1: 2-(4-methyl-2-oxo-2H-chromen-7-yloxy)-N'-(1-(4-aminophenyl)ethylidene)acetohydrazide

#### **Physical features:**

a) Percentage yield: 72%
b) Melting point: 215°C
c) Rf value: 0.32
d) Mobile phase: Ethyl acetate: Hexane (1:1).

Synthesis of 2-(4-methyl-2-oxo-2H-chromen-7-yloxy)-N'-(1-(2 hydroxy phenyl) ethylidene) acetohydrazide [M2]<sup>32</sup>: A mixture of 2-3 drops of glacial acetic acid in compound 2-(4-methyl-2-oxo-2H-chromen-7-yloxy) acetohydrazide (0.0008 mol), o-hydroxyacetophenone (0.0008 mol) and anhydrous alcohol (60 mL) was refluxed on a water bath for 16 hours. The reaction mixture was poured into crushed ice. Filter, dry and recrystallize from ethanol to give compound (M2).

#### **Physical features:**

a) Percentage yield : 76% b) Melting Point : 264 <sup>0</sup>C c) Rf value : 0.24 d) Mobile Phase : Ethyl acetate: Methanol (1:1)



Scheme 2: 2-(4-methyl-2-oxo-2H-chromen-7-yloxy)-N'-(1-(2-hydroxyphenyl)ethylidene)acetohydrazide

#### Antioxidant activity

**DPPH Assay:** The radical scavenging ability of synthesized compounds and the ascorbic acid (standard) was tested on the basis of radical scavenging effect on a DPPH free radical. Different concentrations (20, 50, 100, 200 and 400 g/mL) of compounds and standard were prepared in methanol. In clean and labeled test tubes, 2mL of DPPH solution (0.002% methanol) was mixed with 2mL of different in concentrations of compounds and standard separately. The tubes were incubated at room temperature in dark for 30 minutes and the optical density was measured at 517nm using UV-Visible Spectrophotometer. The absorbance of the DPPH control was also noted<sup>33,34</sup>. The scavenging activity was calculated using the formula: Scavenging activity (%) =(absorbance of control — absorbance of sample / absorbance of control)  $\times$  100.

#### **Results and Discussion**

Two 7-hydroxy 4-methylcoumarin derivatives were prepared according to the scheme. Yields for all compounds were calculated. Their physical constants and thin layer chromatography confirmed the purity of the synthesized compounds. The structure of the synthesized compound was confirmed by IR and 1H NMR spectroscopy.

The IR spectrum is obtained by preparing KBr pellets with a Shimadzu FTIR spectrophotometer 8300 and is expressed as a wave number in cm<sup>-1</sup>. The 1H NMR spectra of the synthesized compound were recorded in DMSO using an AV300BROKEJEOL on a 300MHz spectrophotometer.



Structure of 2-(4-methyl-2-oxo-2H-chromen-7-yloxy)-N'-(1-(4-aminophenyl)ethylidene)acetohydrazide



Date Created: thu aug 23 11:44:10 2012 India Standard Time (GMT+5:30)

# Figure 1: IR Spectra of 2-(4-methyl-2-oxo-2H-Chromen-7-yloxy)-N-(1-(4-aminophenyl) ethylidene) acetohydrazide(M1)



Table 1 **IR Spectra value of M1** 

Figure 2: NMR spectra of (M1)

NMR spectra value of (M1)							
S.N.	Chemical shift(δ)(ppm)	No. of protons	Inferences				
1.	10.61	1H	-NH				
2.	6.12-7.98	8 H	Aromatic				
3.	5.06	2 H	NH <sub>2</sub>				
4.	5.23	2H	-OCH <sub>2</sub>				
5.	3.32	3Н	CH <sub>3</sub> attached to coumarin ring				
6.	2.23	3Н	CH <sub>3</sub>				

Table 2



Structure of 2-(4-methyl-2-oxo-2H-chromen-7-yloxy)-N'-(1-(2-hydroxyphenyl)ethylidene)acetohydrazide



Date Created: thu aug 23 11:45:18 2012 India Standard Time (GMT+5:30)

Figure 3: IR Spectra of 2-(4-methyl-2-oxo-2H-chromen-7-yloxy)-N-(1-(2 hydroxyphenyl)ethylidene) acetohydrazide(M2)

Table 3IR Spectra value M2						
S.N. v (cm <sup>-1</sup> ) Functional Group Assignme						
1.	3309	N-H str.				
2.	3075	C-H str. Aromatic				
3.	2927	C-H str. Aliphatic				
4.	1696	C=O str.				
5.	1614	C=O str.				
6.	1556	C=C str. aromatic				
7.	1513	C=N str.				
8.	1390	C-N str.				
9.	1081	C-O str. in C-O-C				



Table 4

NMR Spectra value (M2)									
S.N.	Chemical shift(δ)(ppm)	No. of protons	Inferences						
1.	10.72	1H	-OH						
2.	10.41	1 H	-NH						
3.	6.12-8.01	8 H	Aromatic						
4.	5.24	2H	-OCH <sub>2</sub>						
5.	3.34	3H	CH <sub>3</sub> attached to coumarin ring						
6.	2.41	3H	CH <sub>3</sub>						

 Table 5

 Scavenging activity of different concentration in %

S.N.	Scavenging activity of different concentration (µg/ml)						
	20	50	100	200	400		
Control	79.34	82.54	89.21	95.45	97.54		
Test (M1)	82.98	89.39	85.42	92.87	95.75		
Test (M2)	83.65	81.47	91.45	96.07	99.04		

Antioxidant Activity: The antioxidant activity at different concentrations 20, 50, 100, 200 and 400  $\mu$ g/ml of the synthesized compound and ascorbic acid was tested on the basis of the radical scavenging effect of the stable DPPH free radical assay. The obtained results were recorded in table 5 In this study, the absorbance was found to increase with the dose of compound and standard.

#### Conclusion

Coumarin derivatives play an important role for several of biological activities. We synthesised two coumarin derivatives M1 and M2 characterised by chromatography, melting point, FTIR and NMR. The synthesised compounds were screened for antioxidant activity.

#### Acknowledgement

The authors would like to thank the authorities of Quantum University for providing the laboratory facilities and support for this work.

#### References

1. Abramov R.S. and Bankoort D.L., *Journal of Physical Chemistry*, **31(1)**, 23 (**1993**)

2. Al-Amiery A.A., Al-Bayati R., Saour K. and Radi M., Cytotoxicity, Antioxidant and Antimicrobial activities of novel 2quinolone derivatives derived from coumarins, *Research on Chemical Intermediates*, **156**, 654-675 (**2011**)

3. Arora R.K., Kaur N., Bansal Y. and Bansal G., Novel coumarin– benzimidazole derivatives as antioxidants and safer antiinflammatory agents, *Acta Pharmaceutica Sinica B*, **4**(**5**), 368–375 (**2014**)

4. Arti R.V., Vijayakumar M., Chandana V.R. and Chandra S.M., In vitro and in vivo antioxidant properties and DNA damage protective activity of green fruit of Ficusglomerata, *Food Chem. Toxicol.*, **48**, 704–709 (**2010**)

5. Aruoma O.I. and Cuppett S.L., Antioxidant Methodology: In Vivo and In Vitro Concepts; AOCS Press: Champaign, IL, USA, 141–172 (**1997**)

6. Azizian J., Mohammadi A., Bidar I. and Mirazaei P., KAl(SO4)2·12H2O (alum) a reusable catalyst for the synthesis of some 4-substituted coumarins *via* Pechmann reaction under solvent-free conditions, *Montash. Chem.*, **139**, 805–808 (**2008**)

7. Brafola G., Fringuelli F., Piermatti O. and Pizzo F., Simple and efficient one-pot preparation 3- substituted coumarin in water, *E-International Scientific Journal of Chemistry*, **43**, 1257 (**1996**)

8. Campbell H.A. and Link K., Studies on the hemorrhagic sweet clover disease, II, The bioassay of hemorrhagic concentrates by following the prothrombin level in the plasma of rabbit blood, *Journal of Biological Society*, **21**, 138 (**1941**)

9. Do Nascimento J.S., Conceição J.C.S. and De Oliveira Silva E., Biotransformation of Coumarins by Filamentous Fungi: An Alternative Way for Achievement of Bioactive Analogs, *Mini. Rev. Org. Chem.*, **16**, 568–577 (**2019**)

10. El-Saghier A. et al, New synthetic approaches to condensed and spiro coumarins: Coumarin-3-thiocarboxamide as building block for the the synthesis of condense and spiro coumarins, *Phosphorus Sulfur Silicon*, **160**, 105–119 (**2000**)

11. Furniss B.S. et al, Vogel's Text book of Practical Organic Chemistry, 5 ed., Longman Scientific and Technical, England, 1163 (**1989**)

12. Garazd M.M., Muzychka O.V., Voyk A.I., Nagorichna I.V. and Ogorodniichuk A.S., Modified coumarins, 27 Synthesis and antioxidant activity of 3-substituted 5,7-dihydroxy-4-methylcoumarins, *Chem. Nat. Compd.*, **43**, 19–23 (**2007**)

13. Heravi M., Sadjadi S., Oskooie H., Shoar R. and Bamoharram F., The synthesis of coumarin-3-carboxylic acids and 3-acetyl-

coumarin derivatives using heteropolyacids as heterogeneous and recyclable catalysts, *Catal. Commun*, **9**, 470–474 (**2008**)

14. Hoult J.R. and Paya M., Pharmacological and biochemical actions of simple coumarins: natural products with therapeutic potential, *Gen. Pharmacol*, **27(2)**, 713-22 (**1996**)

15. Inoue Y., Kondo H., Taguchi M., Jinbo Y., Sakamoto F. and Tsukamoto G., Synthesis and Antibacterial Activity of Thiazolopyrazine-Incorporated Tetracyclic Quinolone Antibacterials, *Journal of Medicinal Chemistry*, **37**, 586 (**1994**)

16. Kadhum A.A.H. et al, The Antioxidant Activity of New Coumarin Derivatives, *Int. J. Mol. Sci.*, **12**, 5748-5758 (**2011**)

17. Kathuria A., Gupta A., Priya N., Singh P., Raj H.G., Prasad A.R., Parmar V.S. and Sharma S.K., *Bioorganic Medicinal Chemistry*, **17**(4), 1550 (**2009**)

18. Kennedy R.O. and Thornes R.D., Coumarins: Biology, Applications and Mode of Action; John Wiley and Sons: Chichester, England (**1997**)

19. Kotali A., Lafazanis I. and Harris P., Synthesis of 6,7diacylcoumarins *via* the transformation of a hydroxy into a carbonyl group, *Synth. Commun.*, **38**, 3996–4006 (**2008**)

20. Mazumder A., Wang S., Neamati N., Nicklaus M., Sunder S., Chen J., Milne G., Rice W., Burke T. and Pommler Y., Antiretroviral Agents as Inhibitors of both Human Immunodeficiency Virus Type 1 Integrase and Protease, *Journal* of Medicinal Chemistry, **39**, 2472 (**1996**)

21. Murray R.D.H., Coumarins, Nat. Prod. Rep., 6, 477–505 (1995)

22. Musicki B., Periers A.M., Laurin P., Ferroud D., Benedetti Y., Lachaud S., Chatreaux F., Haesslein J.L., Iltis A. and Pierre C., Improved antibacterial activities of coumarin antibiotics bearing 5',5'-dialkylnoviose: biological activity of RU79115, *Bioorganic Medicinal Chemistry*, **10**, 1695 (**2000**)

23. Nofal Z.M., El-Zahar M. and Abd El-Karim S., Novel coumarin derivatives with expected biological activity, *Molecules*, **5**, 99–113 (**2000**)

24. Noroozi-Pesyan N., Khalaf J. and Malekpoor Z., Pro Color Colorants Coat, 2, 61-70 (2009)

25. Penta S., Introduction to Coumarin and SAR, In Advances in Structure and Activity Relationship of Coumarin Derivatives, Academic Press, Cambridge, MA, USA, 1–8 (**2016**)

26. Phutdhawong W. et al, Synthesis and Biological Activity Evaluation of Coumarin-3-Carboxamide Derivatives, *Molecules*, **26**, 1653 (**2021**)

27. Rajitha B., Kumar V., Naveen Someshwar P., Venu Madhav J., Reddy P.N. and Reddy Y., Thirupathi, *Online Journal of Organic Chemistry*, **12(2)**, 23-27 (**2006**)

28. Saini S., Neerupma N., Mittal A. and Kumar G., synthesis and antioxidant activity of the 2-methyl benzimidazole, *Journal of Drug Delivery and Therapeutics*, **6(3)**, 100-102 (**2006**)

29. Satyanarayan V.S., Sreevani P. and Sivakumar A., Synthesis and antimicrobial activity of new Schiff bases containing coumarin moiety and their spectral characterization, *Arkivoc*, **17**, 221–233 (**2008**)

30. Singh A.D., Kumar S., Saini S. and Prabha H., Synthesis and biological evaluation some new of 7-hydroxy-4-methylcoumarin derivative, *APJHS*, **4**(4), 84-94 (2017)

31. Smitha G. and Sanjeeva R., ZrCl4-catalyzed Pechmann reaction: Synthesis of coumarins under solvent-free conditions, *Synth. Commun.*, **34**, 3997–4003 (**2004**)

32. Takadate A., Masuda T., Murata C., Isobe A., Shinohara T., Irikura M. and Goya S., A Derivatizing Reagent-Kit Using a Single

Coumarin Fluorophore. Analytical Sciences, *Analytical Sciences*, **13**, 753 (**1997**)

33. Xu J. et al, Cytosporones, coumarins and an alkaloid from the endophytic fungus Pestalotiopsis sp. isolated from the Chinese mangrove plant Rhizophoramucronata, *Bioorg. Med. Chem.*, **17**, 7362–7367 (**2009**)

34. Zabradnik M., The Production and Application of Fluorescent Brightening Agents, John Wiley and Sons: New York, NY, USA (1992).

(Received 02<sup>nd</sup> January 2022, accepted 01<sup>st</sup> March 2022)